



Evaluation of Hepatocarcinogenicity Biomarkers in 3D HepG2 Liver Spheroids Following Nanomaterial Exposure

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Introduction:

- Due to the rapid development and implementation of a diverse array of engineered nanomaterials (ENM), exposure to ENM is inevitable and the development of robust, predictive *in vitro* test systems for hazard characterisation is essential.
- Adverse Outcome Pathways (AOP) describe the sequence of biological events known as key events (KEs) that are required to result in a pathological event and are therefore considered to be useful mechanistic tools for the development of novel endpoint targets for human and environmental risk assessment.
- Understanding AOPs permits the identification of possible mechanistic biomarkers that have the potential to be integrated into advanced *in vitro* testing systems, to improve predictive toxicology.

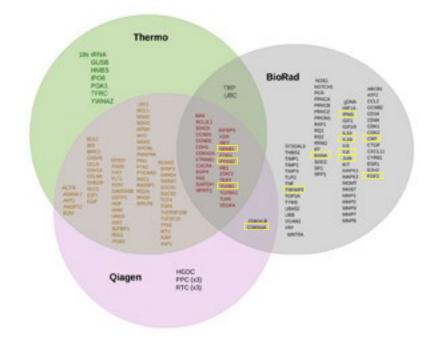


Figure 1. PTGS to Hepatocellular Carcinoma Gene Array Comparison. A cross gene analysis was performed of the genes on pre-defined liver cancer PCR array plates from three companies (Qiagen, Thermo Scientific, and Bio-Rad). Pathway based similarity analysis confirmed that the Bio-Rad plate demonstrated closer overlap (17s genes) with predictive toxicogenomics space (PTGS) liver specific components (which cover 299 genes) when compared to the Thermo Scientific and Qiagen plates (10 and 9 genes respectively)

Aim:

Table 1. A summary of the key genes listed throughout

all Liver AOP specific key events. All genes in red are included in the Bio-Rad Hepatocarcinoma PCR Array.

NFKB2

NFKBIA

NFKBIE

PTGS2

PDGFA

RXRA

SERPINE1

SMAD3

SMURF2

TGFB2 - TGFB1

Key Genes listed throughout all Liver AOP specific key events

ADM

CDKN1A

CEBPB

CFLAR

CXCL3 FGF2

ICAM1

IER3

IFNG

IL1B

CXCL2 – CXCL12

The aim of this study was to develop a novel panel of biomarkers to detect KEs or Molecular Initiating Events (MIEs) that are indicative of AOP. This will allow us to develop the current understanding of human liver AOPs in relation to ENM exposure scenarios and to develop a more targeted *in vitro* testing approach to predict adverse outcomes.

Methods:

- A cross gene analysis was performed of different PCR arrays and genes associated with Liver AOP specific key events (figure 1 & table 1).
- HepG2 3D liver spheroids (figure 2) were used as an effective screening approach for adverse effects to human health following exposure to ENM. The 3D HepG2 cell line based liver model was developed to support both short and long term ENM exposure regimes.
- 3D HepG2 liver spheroids were exposed to either TiO₂ or Ag for up to 120 hours prior to RNA being extracted (figure 3).
- Reverse transcription real-time polymerase chain reaction (RT-qPCR), using a predefined hepatocellular carcinoma PCR array, was used to allow for the study of multiple molecular pathways at a time.

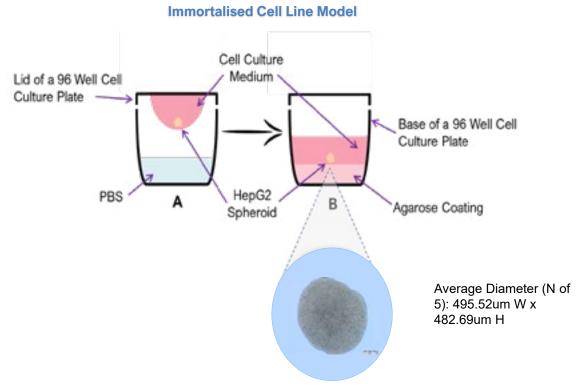


Figure 2. Schematic diagram displaying the formation of 3D HepG2 spheroid model using the hanging drop method initially (A) before transferring into agarose coated well plates (B).

Table 3: Changes in the 3D liver spheroids gene expression profile following 120hr exposure to AFB1, Ag and TiO₂. Red cells indicate upregulation and

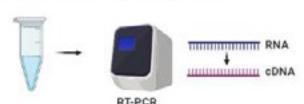
1 Extract RNA
3D HepG2 spheroids

RT-qPCR Array Workflow





3 Reverse transcribe RNA to cDNA







Gene Expression analysis

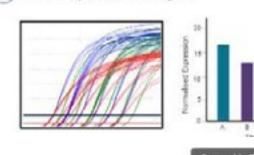


Figure 3: Schematic Diagram of RT-qPCR Workflow (Created with BioRender.com).

Table 4. Key genes highlighted from gene expression qPCR

arrays under differe	ent treatment condition	ons.
24 Hours	Ag	TiO ₂
	RXRA	IGFBP3
	1	TGFB1
	1	WNT5A
120 Hours	Ag	TiO ₂
	IL10	IL10
	CD44	1
	CDNK1A	1
	ESR1	ESR1
	MMP3	MMP3
	TGFBR2	TGFBR2

Results:

Table 2: Changes in the 3D liver spheroids gene expression profile following 24hr exposure to AFB1, Ag and TiO₂. Red cells indicate upregulation & yellow cells indicate down regulation. *FC, fold change, SEM, standard error of the mean*.

	AFB1		AG 1.0		AG 5.0		TiO2 1.0		TiO2 5.0	
	FC	SEM	FC	SEM	FC	SEM		SEM		SEM
ATF2					7.188	18.214				
BCL2L1					4.671	15.081				
CCND2	19.356	16.725			8.109	26.991				
CD44					21.927	54.750				
CDH1					4.244	12.176				
CDKN1A					20.500	51.187				
CDKN2A					6.707	13.194				
CTNNB1					4.667	15.179				
GF2					15.692	39.182				
HIF1A					7.720	16.937				
GF1							10.231	12.594		
GFBP3					5.307	15.228	4.786	4.088	6.213	2.99
L8					6.878	22.261				
IUN					7.613	20.998				
KIT			5.532	2.061	24.170	60.349				
MAPK1					7.027	17.459				
мет					6.111	17.388				
MMP1					11.189	27.938				
ММР2	-4.572	0.189			24.392	60.904				
ММР7					4.198	10.482	-4.882	0.072		
NFKB1					7.738	22.703				
NOTCH1	6.225	5.379								
PRKCB					9.437	23.563				
PTEN					20.385	48.127				
RRM2					4.676	8.399				
RXRA					-4.415	0.970				
SP1					8.443	20.487				
TGFB1					4.393	17.217	5.069	3.818	4.264	3.59
THBS1					4.019	11.357				
VCAM1	5.673	4.902			8.665	21.635				
WNT5A	4.491	3.880			11.550	25.678	5.923	2.518		

TiO₂ 1.0

■ TiO₂ 5.0

Acute (24 Hour) Exposure

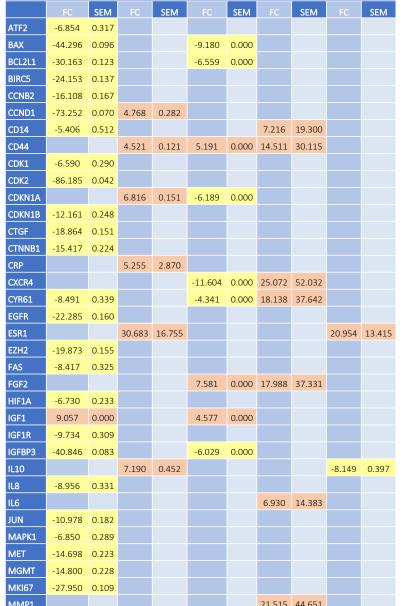
WNT5A

AFB1 TiO, 1.0

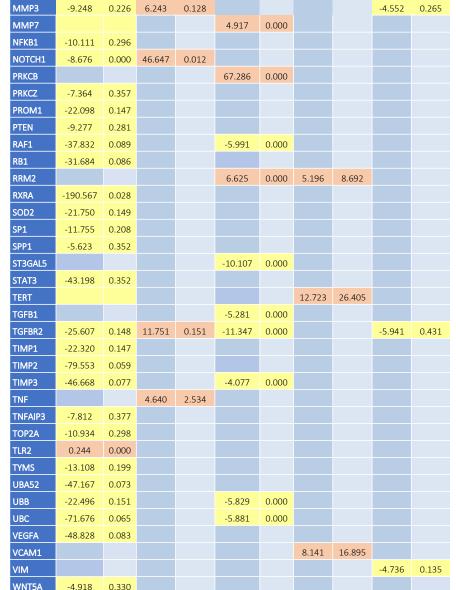
Figure 4: Changes in gene expression of IGFBP3, TGFB1

(a) and WNT5A (b) following exposure to TiO2 and AFB1 for

24 hours. Mean data \pm SEM presented n = 3.



yellow cells indicate down regulation. FC, fold change, SEM, standard error of the mean



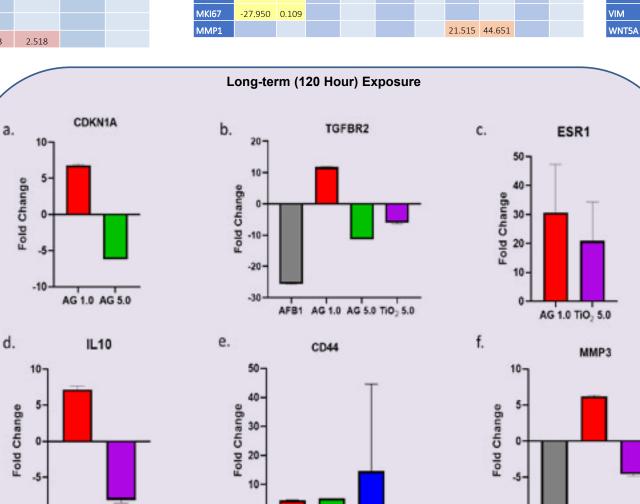


Figure 5: Alterations to (a) CDKN1A, (b) TGFBR2, (c) ESR1, (d) IL10, (e) CD44, (f) MMP3 expression following exposure to Ag, TiO₂ (1.0 & 5.0 μg/ml) and AFB1 (positive control) for 120 hours. Mean data ± SEM presented n = 3.

AG 1.0 AG 5.0 TiO, 1.0

Summary of key findings:

- Exposure of Ag and TiO₂ to 3D HepG2 liver spheroids results in transcriptional alternations in genes known to be important in driving hepatocellular carcinoma. These changes appear to differ depending on the whether it is an acute or chronic exposure period (table 2 & 3).
- Key genes from each time point following the PCR array analysis have been highlighted (figure 4 & 5). Interestingly, the genes highlighted after 24hr exposures have been linked to both liver fibrosis and inflammation which known are precursors liver carcinogenesis.
- Due to little overlap between genes highlighted at 24 and 120hr, the transcriptional changes are time specific (table 4). At 24hr, shorter term changes are most likely related to early stage changes such as inflammation and fibrosis; while longer exposures are associated with transcriptional changes that are more relevant in carcinogenesis.
- By using the qPCR liver cancer arrays, we have been able to highlight a number of genes (table 4) that should be explored further to better understand their role in liver associated carcinogenesis following long-term ENM exposure.





AFB1 AG 1.0 TiO, 5.0





AG 1.0 TiO2 5.0

Acknowledgments: