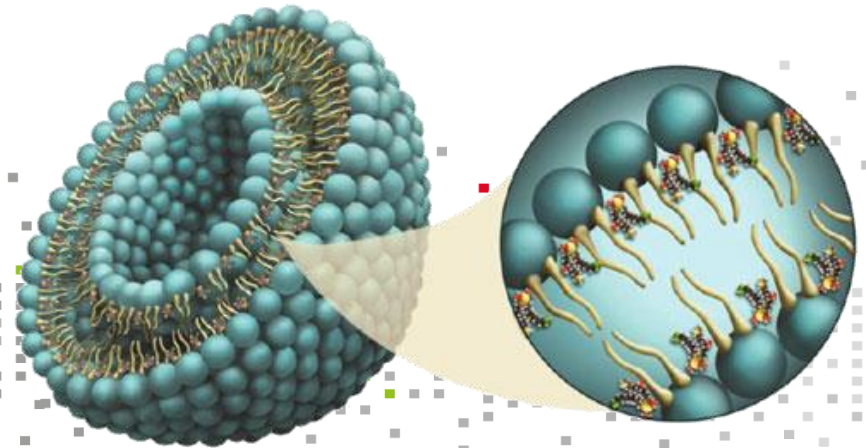


liten
cea tech



CHARACTERIZATION OF NANO CARRIERS FOR DRUG DELIVERY SYSTEMS: THE LIPIDOTS®

NanoSafe 2016 | Amandine Arnould | 07-11/11/2016



Nano-safety issue

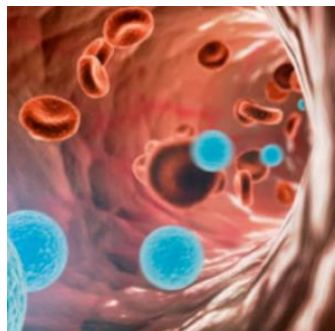
Environment



- Air decontamination
- Water treatment

Fe, Al

Medicine



- Bactericide
- Drug carriers

Ag, Liposomes

Agribusiness



- Food colouring
- Anticaking

TiO₂, SiO₂

Cosmetic

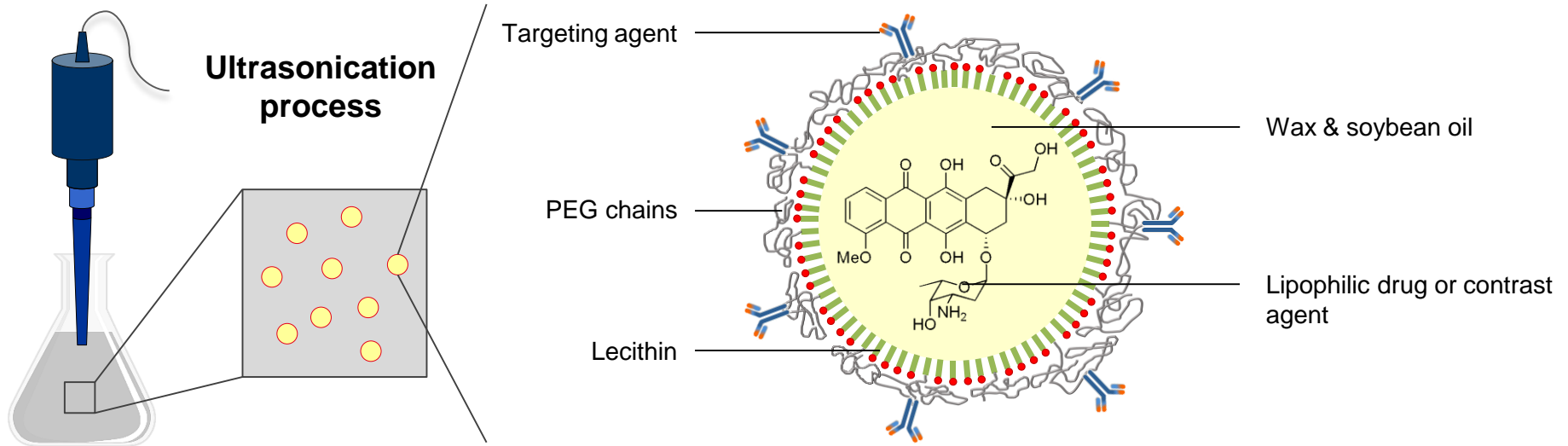


- UV filter
- Colouring

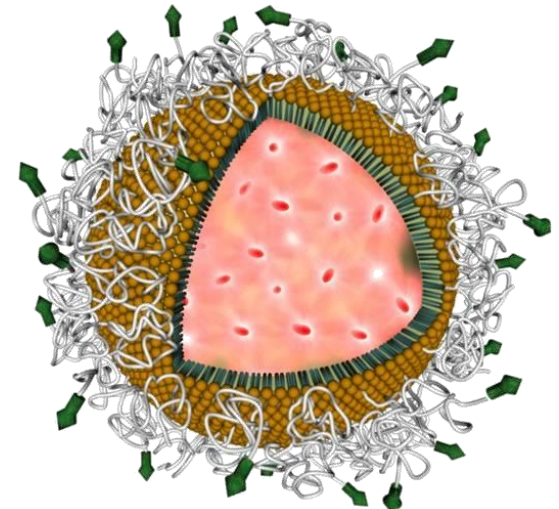
Core shell TiO₂

Robust characterization on organic nanoparticles

- Stability of the nanoparticles (aging)
- Interaction with biological media (toxicity)



- Tunable diameter (lipid ratio) (30 to 120 nm)
- Completely biodegradable & biocompatible
- **Lipophilic** drug or contrast agent
- Amorphous particles



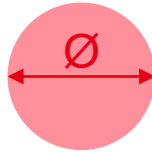
EQUIPEX



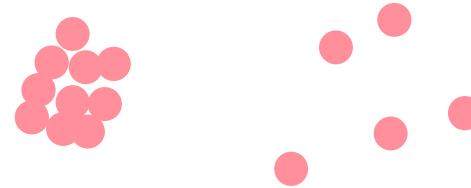
Transmission Electron Microscopy



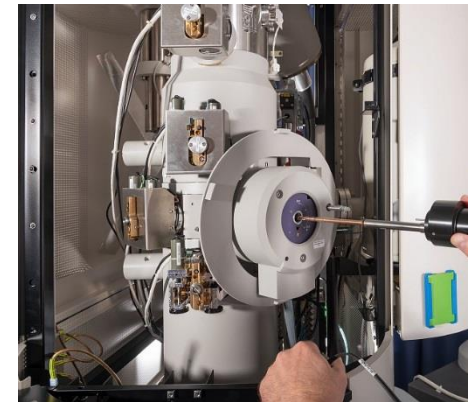
Size



Agglomeration, dispersion



- Drying process
- Negative staining + drying process
- Plunge freezing (cryo-TEM)
- In-situ liquid



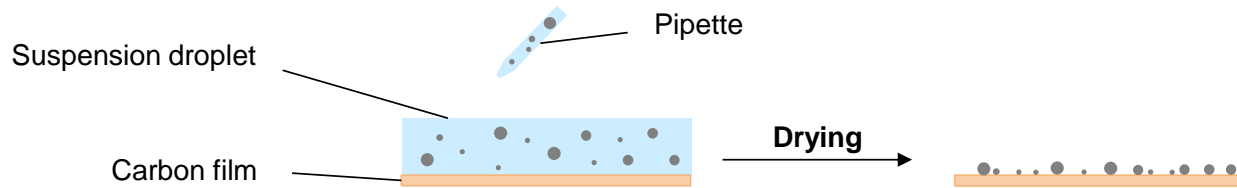
➡ Well adapted for metallic particles

➡ Optimizations for organic particles

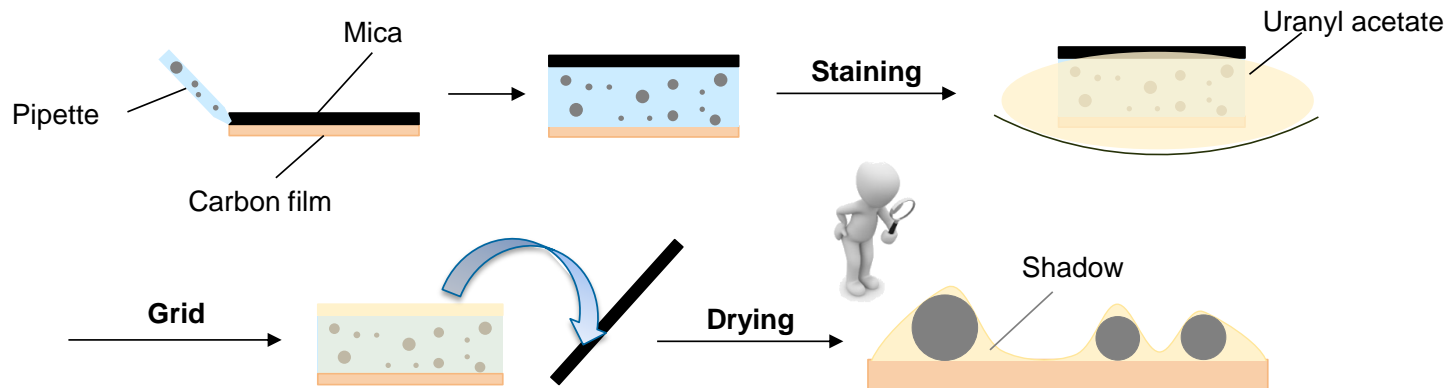


Drying process: concentration adjustment

■ Without staining:

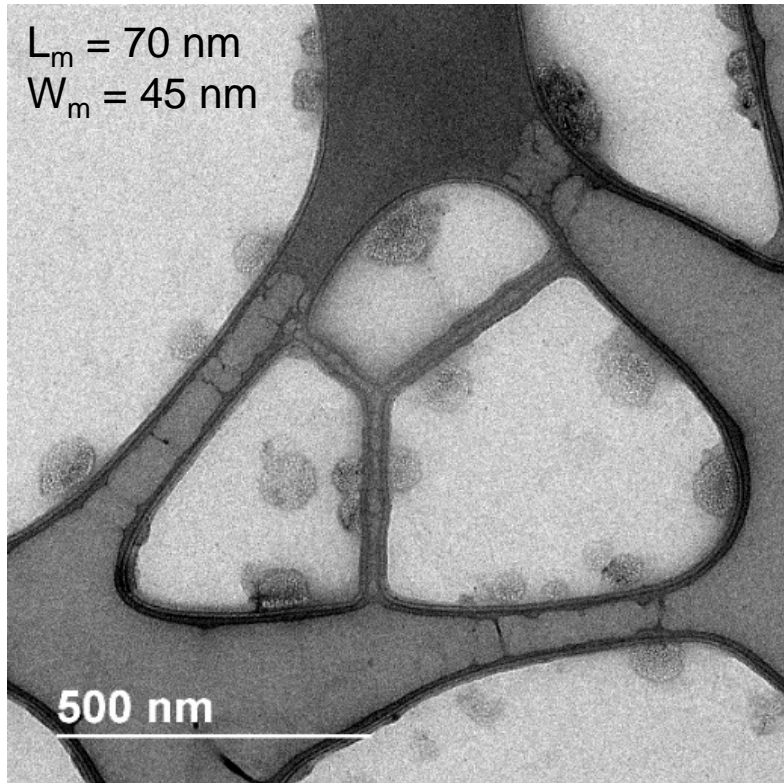


■ With negative staining:



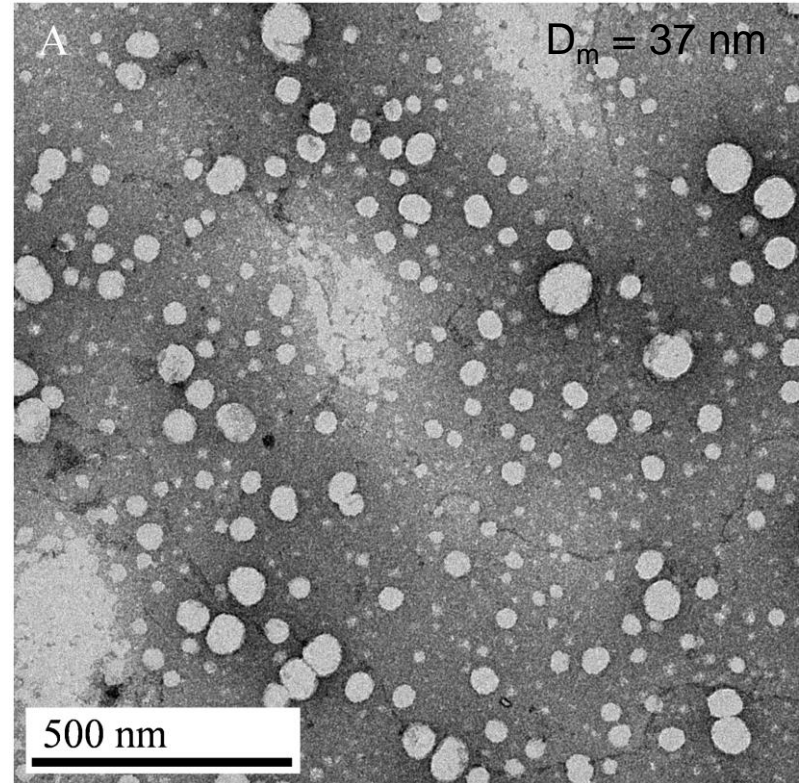
➔ Quick methods

Without staining



- Particle dehydration
- Poor contrast
- Concentration on the lacey carbon

With negative staining

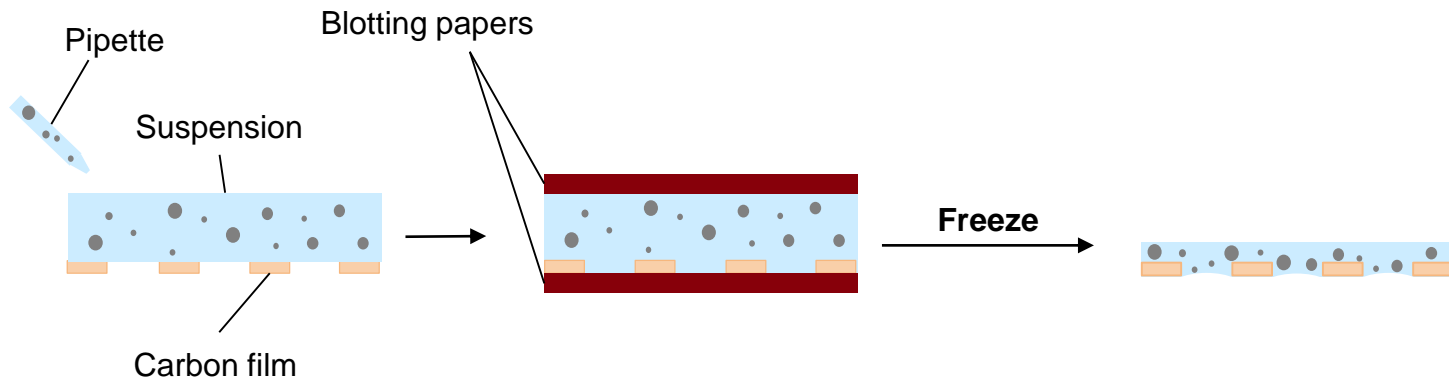
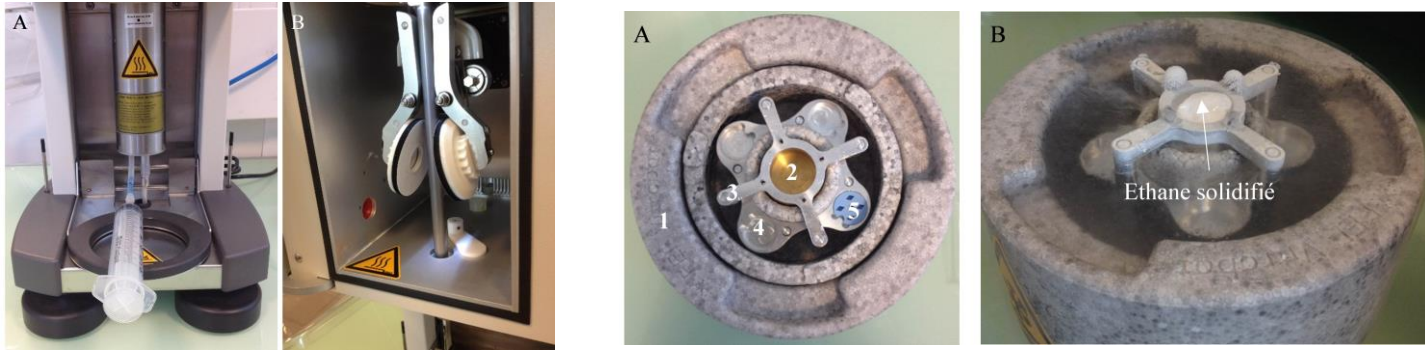


- Real shape or artefact ?
- Agglomeration
- Interior of the particle not visible

➡ Not appropriate



Cryogenics: preservation and 3D reconstruction

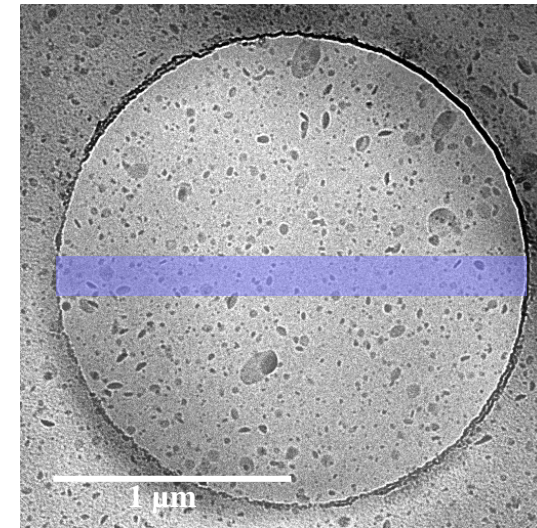
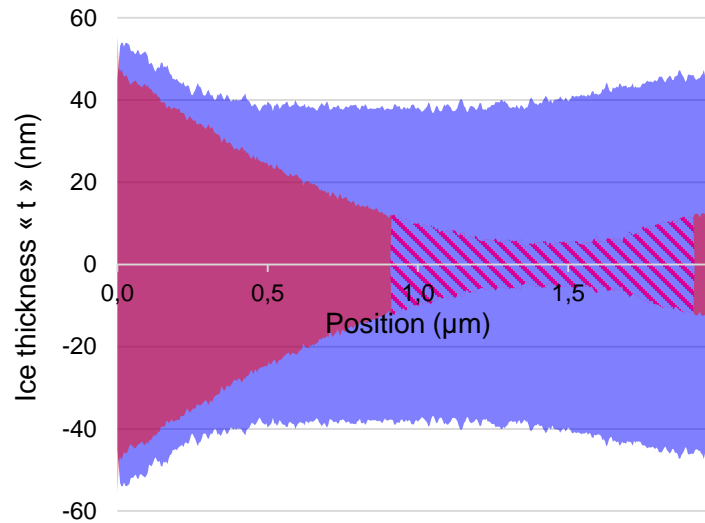
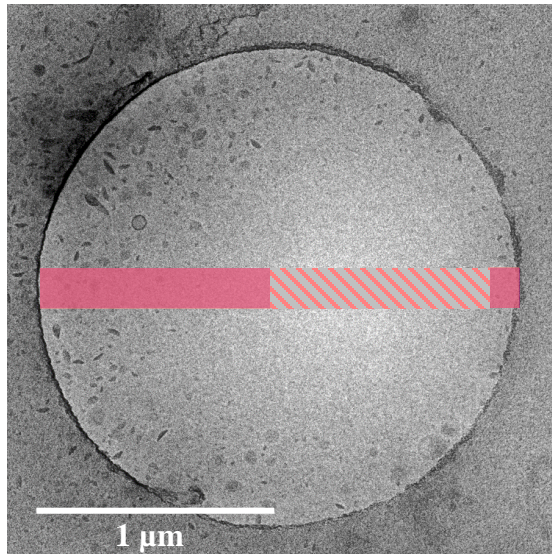


➔ Amorphous & homogenous ice: optimizations

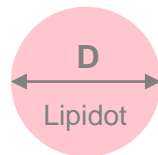
Cryogenics: ice thickness optimizations

Blot: 3s

Blot: 1s



- $t < D_{\min}$
- $D_{\min} < t < D_{\max}$

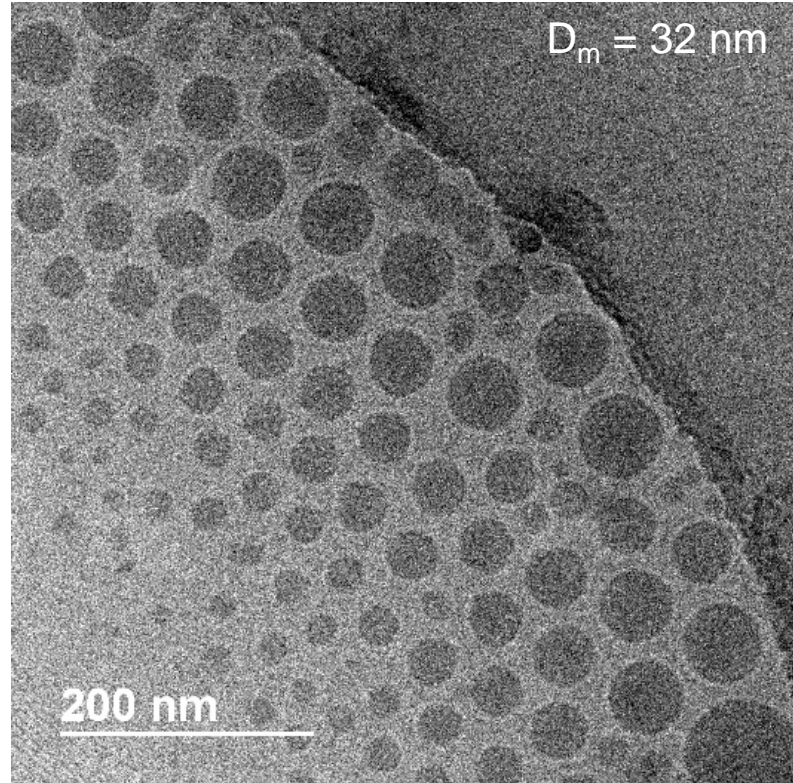
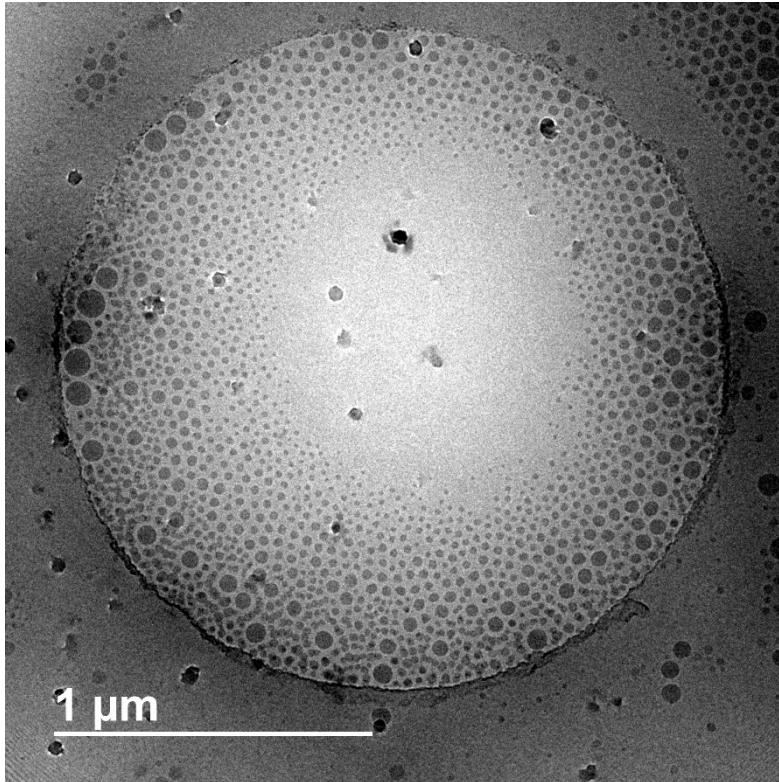


$$20 \text{ nm} < D < 80 \text{ nm}$$

$D_{\min} < t < D_{\max}$

Homogenous repartition of the particles on the grid

TEM OBSERVATIONS

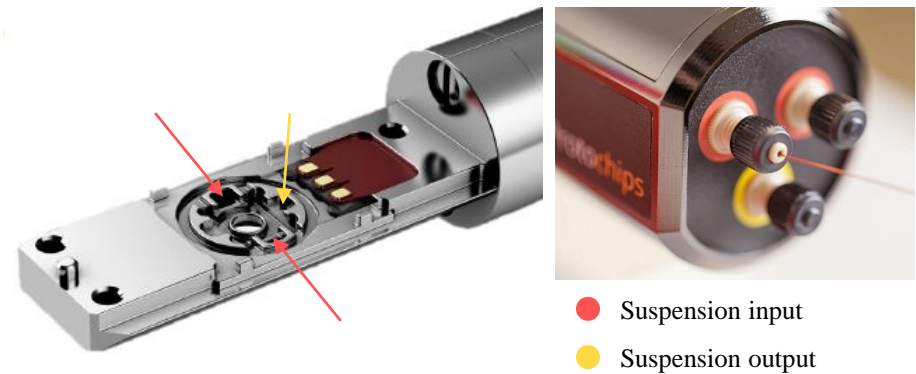
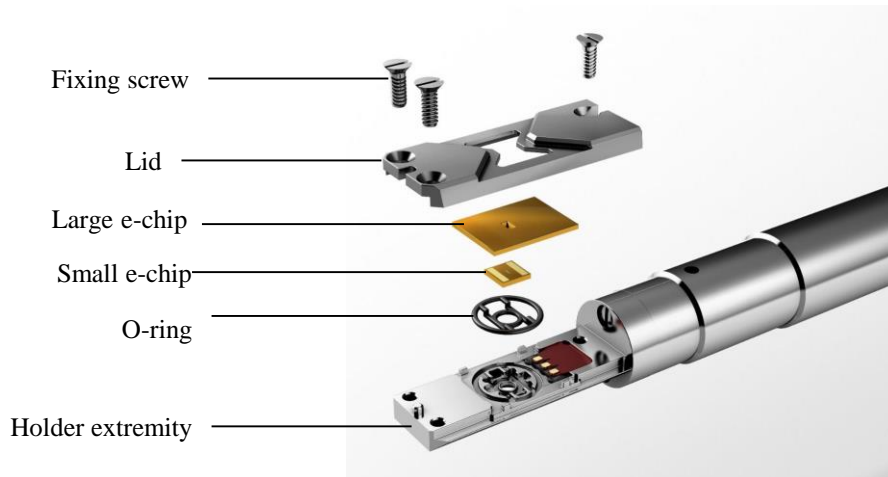
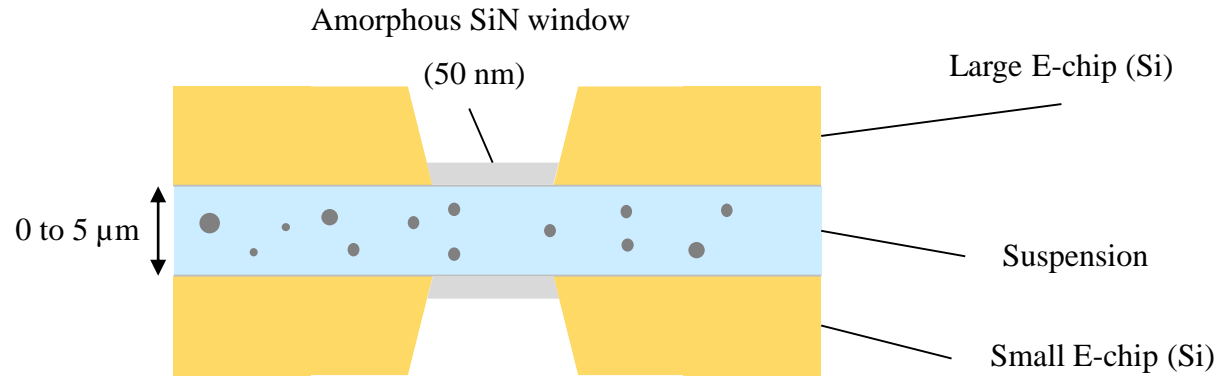


- Quite good contrast
- Ice contaminations
- Round shape

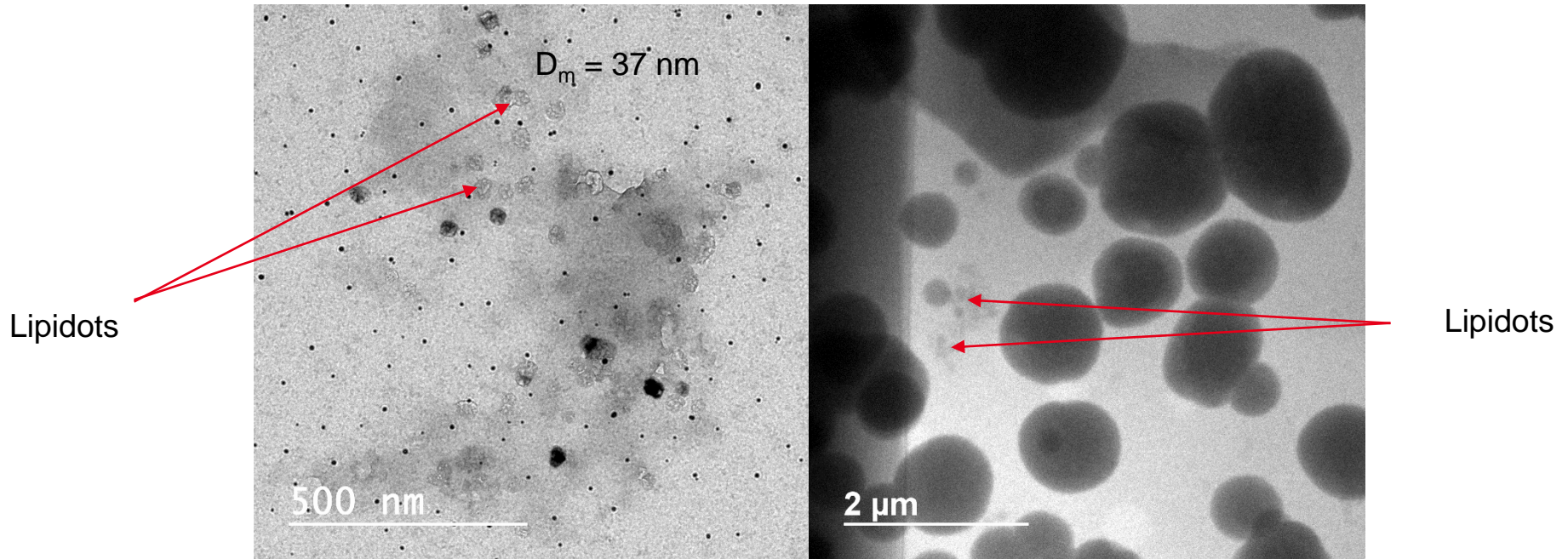
➡ Native state cryofixed

TEM OBSERVATIONS

In-situ liquid: native state check-in for particles in suspension



TEM OBSERVATIONS



- Contaminations
- Poor contrast
- Lack of statistic

➡ Clean the holder & retry with smaller spacer (50 nm)

CONCLUSION

Comparison of the different sample preparation methods

■ Drying process

poor contrast

no special equipment required

quick

■ Negative staining + drying

Preserved shape

dehydration

$$L_m = 70 \text{ nm}$$

$$W_m = 45 \text{ nm}$$

$$D_m = 37 \text{ nm}$$

overstatement of the diameter

■ DLS

$$D_h = 71,8 \text{ nm}$$

flattening

concentration on the lacey carbon

Interior of the particle non visible

agglomeration

■ Plunge freezing

lots of particles

$$D_m = 32 \text{ nm}$$

■ In-situ liquid

no preparation artefact

good contrast

$$D_m = 37 \text{ nm}$$

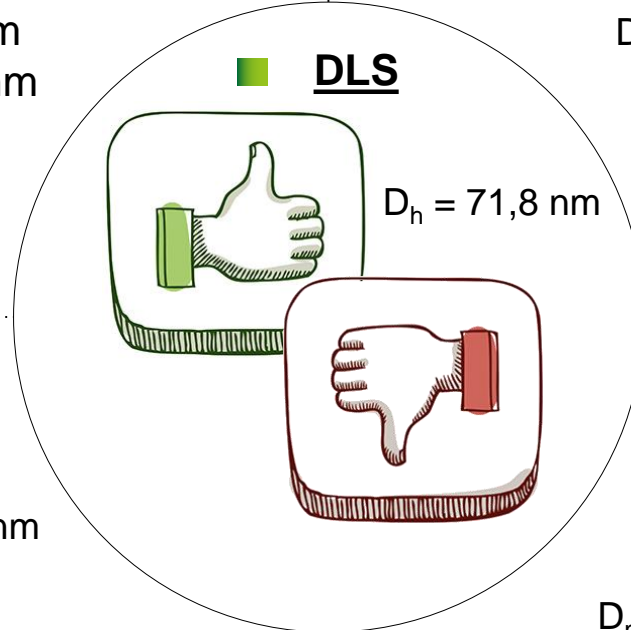
poor contrast

ice thickness

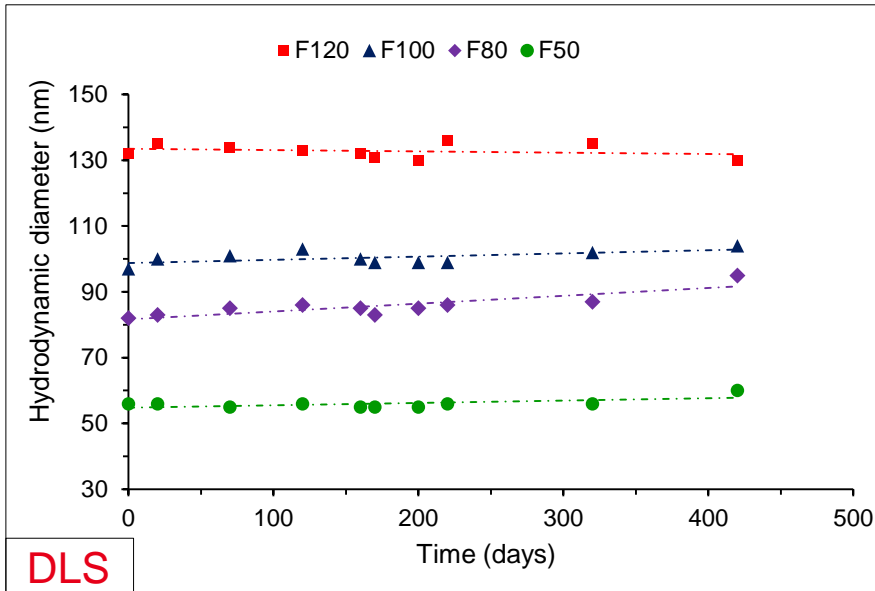
ice contamination

native state

time consuming



■ Stability of reference particles in PBS



NC75 (75% wax, 25% soybean oil), 4°C

● High stability (> 13 months)

PBS (*Phosphate Buffered Saline*): Buffer solution

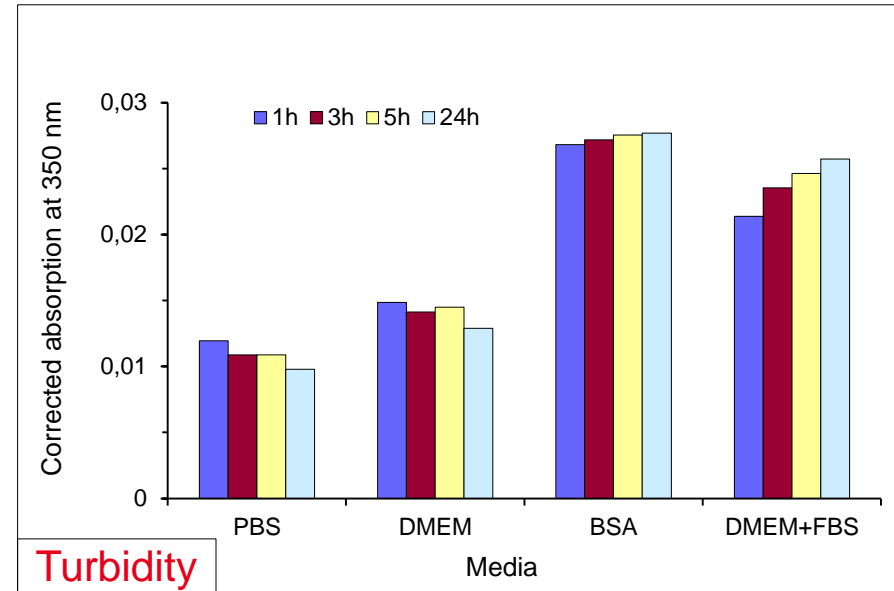
DMEM (*Dulbecco's Modified Eagle Medium*): ~ plasma without protein

BSA (*Bovine Serum Albumin*): ~ liver proteins

FBS (*Foetal Bovine Serum*): ~ plasma with plasma proteins & biomolecules

[1]

■ Stability in biological media



F50 NC75, 0,2 % m/m, 37°C

● DMEM ↔ PBS ; stable over 24h

● BSA: quite stable over 24h ; increased of the particle diameter compared to PBS ?

● DMEM + FBS: destabilization of the particles ?
Adsorption of proteins at the surface of the particles ?



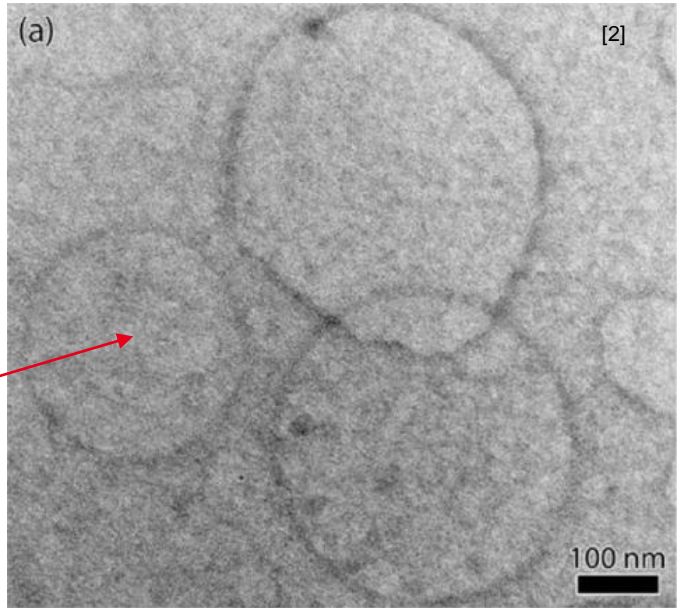
Thank you for your attention



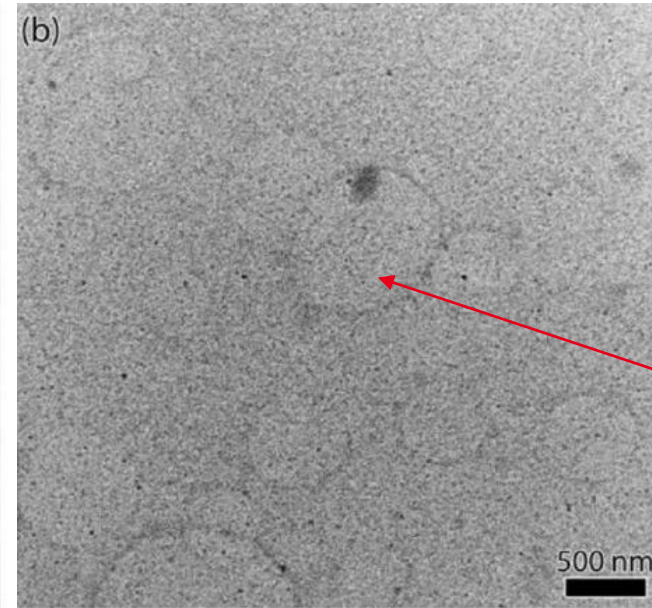
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- Fanny Caputo (CEA)
- Anne-Claude Couffin (CEA)
- Isabelle Texier-Nogues (CEA)
- Constantin Mattei (CEA)
- Benoit Gallet (IBS)
- Jean-François Damlencourt (CEA)
- Romain Soulas (CEA)
- Delphine Boutry (CEA)
- Stéphane Aguy (Eden Instrument)



TEM OBSERVATIONS



Liposome



Liposome

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